Sex Pheromones of the Hair Crab *Erimacrus isenbeckii*. II. Synthesis of Ceramides

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To confirm their structures and to assess the pheromonal activity, novel ceramides, possible sex pheromones of the hair crab *Erimacrus isenbeckii*, were synthesized from D-galactose. The synthetic ceramides were identical with the natural ceramides.

The hair crab Erimacrus isenbeckii, a commercially important species, exhibits a series of typical mating behaviors: precopulatory guarding (male grasps female), molting of female, copulation, and postcopulatory guard. The involvement of a pheromone released from pre- and postmolt females in these behaviors was shown by "sponge assay". A sexually competent male exhibits the typical mating behaviors toward a bath sponge containing water conditioned with pre- or postmolt females. Bioassay-guided isolation furnished a mixture of novel ceramides as possible sex pheromones.¹ It is well known that either pheromone structures or the combination of components are crucial for pheromonal activity in insect pheromones.² To confirm the structures proposed for the sex pheromones and to assess pheromonal activity of this ceramide mixture, we attempted to synthesize all ceramides. This paper deals with total synthesis of the ceramides.

Results and Discussion

There are many reports of syntheses of ceramides (cerebrosides);3-7 syntheses of ceramides consisting of 2-hydroxy fatty acids and phytosphingosines have also been established. Usually, (2S,3S,4R)-phytosphingosines were synthesized from natural chiral pools such as D-galactose, 9,10 D-xylose, 11 L-ascorbic acid, 12 glyceraldehyde, 13 and L-serine.¹⁴ The present work employed a protocol of Akiyama and Natori¹⁵ starting from D-galactose, because of its advantage in large-scale synthesis. On the other hand, racemic synthesis of the fatty acid portion starting from 9-bromononanol was envisioned.¹⁶ After the individual synthesis of two sphingosines (C_{15} , C_{16}) and three fatty acids (C₂₃, C₂₄, C₂₅), a combination of these components via mixed condensation was expected to afford a ceramide mixture. Acylation of the phytosphingosines with the racemic fatty acids was expected to give a diastereomeric mixture from which the desired 2'R isomer could be separated. Our synthetic strategy for ceramides is depicted in Scheme 1.

Syntheses of the C₁₅ Phytosphingosine (12a) and C₁₆ Phytosphingosine (12b). Starting from β -D-galactose pentaacetate (2), 3,4,6-tri-*O*-benzyl-D-galactopyranose (6) was prepared in four steps in 54% yield from 2.¹⁷ Oxidation of **6** with sodium metaperiodate provided an aldehyde **7**,¹⁸ which was directly subjected to Wittig reaction without purification; **7** was treated with a C₁₀ phosphonium salt prepared from bromodecane to afford a mixture of geo-

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metrically isomeric alcohols **8a**. Mesylation of **8a** followed by hydrogenation gave rise to a tetraol monomesylate **10a** in 73% yield from **6**. The mesylate **10a** was converted to an azide **11a** with sodium azide in 82% yield. Finally, catalytic hydrogenation of the azide afforded the C_{15} phytosphingosine **12a** in 32% overall yield.

Similarly, C_{16} phytosphingosine was synthesized. The aldehyde **7** and C_{11} phosphonium salt prepared from bromododecane were coupled to give alcohols **8b**. A fourstep reaction sequence from **8b** afforded the C_{16} phytosphingosine **12b** in 23% overall yield from **7**.

Synthesis of C23 Fatty Acid 21a. The isopropyl terminus was initially introduced by treatment of 9-bromo-1-nonanal (14), which was obtained by PCC oxidation of 9-bromo-1-nonanol (13), with the ylide of isobutyltriphenylphosphonium bromide, which furnished a mixture of geometric isomers of branched bromoalkenes 15. The mixture was heated with triphenylphosphine to give phosphonium salts 16. Without purification, 16 was again subjected to a Wittig reaction with 8-bromo-1-octanal. The resulting diene 17a showed complex ¹H NMR signals probably due to olefin migration, which could consequently be converted to a single bromoalkane 18a by hydrogenation. Then, conventional acetamidomalonic ester synthesis was applied to 18a. The resulting mixture was hydrolyzed and decarboxylated to afford racemic amino acid 19a. Deamination of 19a was effected with NaNO₂ in an acidic solution ($H_2SO_4/H_2O/1,4$ -dioxane) to give the 2-hydroxy fatty acid 20a. This was acetylated to afford 21a with Ac₂O and pyridine for the following condensation step (5.7%) overall yield from 13).

Synthesis of C₂₄ (21b) and C₂₅ (21c) Fatty Acids. In a similar manner used for the preparation of the C₂₃ fatty acid 21a, C₂₄ 21b and C₂₅ 21c fatty acids were synthesized using 9-bromononanal or 10-bromodecanal in the second Wittig reaction. Acetates of the C₂₄ and C₂₅ fatty acids were synthesized in 4.9% and 14.5% overall yield, respectively.

Synthesis of the Hair Crab Ceramide Mixture (1a). Prior to the condensation step, each of the sphingosines and fatty acids was mixed according to the natural abundance [**12a**:**12b** (4:5); **21a**:**21b**:**21c** (5:5:3)], which had been roughly estimated from both HPLC and FABMS data of the natural ceramide mixture. Coupling of these components by using WSCI (water-soluble carbodiimide)/HOBt (1-hydroxybenzotriazole) and subsequent deacetylation with K₂CO₃ gave a diastereomeric mixture of ceramides **1a** and **1b**. Silica gel TLC (CHCl₃/MeOH, 95:5) clearly showed two well-separated spots, of which the slowermoving spot (**1a**) corresponded to the natural 2'*R* ceram-





^a Reagents and conditions: (i) HBr/AcOH; (ii) MeOH/TEA; (iii) BnCl/KOH; (iv) H₂SO₄; (v) NaIO₄; (vi) *n*-C₁₀H₂₁PPh₃Br or *n*-C₁₁H₂₃PPh₃Br/*n*-BuLi; (vii) MsCl/pyridine; (viii) H₂/Pd-C; (ix) NaN₃; (x) H₂/Pd-C.

ОН

12b (C₁₆)

8b $R = n - C_{10}H_{21}$

Scheme 3^a

Scheme 2^a



^{*a*} Reagents and conditions: (i) PCC; (ii) *i*-BuPPh₃Br/n-BuLi; (iii) PPh₃; (iv) 8-bromooctanal or 9-bromononanal or 10-bromodecanal/*n*-BuLi; (v) H₂/Pd-C; (vi) (EtO₂C)₂CHNHAc/EtONa, then HCl; (vii) NaNO₂; (viii) Ac₂O/pyridine.

ides. This was then separated by silica gel column chromatography with CHCl₃/MeOH solvent systems.

The synthetic ceramide mixture **1a** was identical with the natural product in the ¹H and ¹³C NMR spectra.¹⁹ In addition, HPLC profiles of **1a** were almost superimposable on those of the natural ceramide mixture, except for the absence of minor components. However, no mating behavior of male crabs could be observed for the synthetic ceramide mixture, because no sexually competent males were available.

Experimental Section

General Experimental Procedures. NMR spectra were recorded on either a Bruker AC-300 (300 MHz for ¹H, 75 MHz





1a m= 1, 2 n= 1, 2, 3

^a Reagents and conditions: (i) WSCI/HOBt; (ii) K₂CO₃; (iii) Si gel column separation.

for ¹³C), a JEOL JMN-α600 (600 MHz for ¹H, 150 MHz for 13 C), or a JEOL JMN- α 500 (500 MHz for 1 H, 125 MHz for 13 C) NMR spectrometer at 303 K. ¹H chemical shifts were referenced to residual solvent peaks: δ 7.24 for CHCl₃ in CDCl₃; δ 3.30 for CD₂HOD in CD₃OD; δ 3.30 for CD₂HOD in CDCl₃/ CD₃OD (1:1). ¹³C chemical shifts were referenced to ¹³C signals of CDCl₃ (δ 77.0) or CD₃OD (δ 49.0). FAB mass spectra were measured on a JEOL JMX-SX102/SX102 tandem mass spectrometer. Glycerol or *m*-nitrobenzyl alcohol (NBA) was used as a matrix. Air- and moisture-sensitive reactions were carried out under an atmosphere of argon. When necessary, solvents were distilled from dehydrating agents (THF from sodium and benzophenone; EtOH from sodium and diethylphthalate; CH2-Cl₂, toluene, MeOH, and 1,2-dichloroethane from CaH₂). Routine monitoring of reactions and preparative TLC were performed using precoated Si gel TLC plates (Merck 60 F₂₅₄). Spots were visualized with either H₂SO₄, anisaldehyde-H₂SO₄, or ninhydrin reagent. Column chromatography was performed on Merck Si gel 60 (60-230 mesh).

3,4,6-Tri-O-acetyl-1,2-O-(1-methoxyethylidene)-a-D-ga**lactopyranose (4)**. To penta-O-acetyl- β -D-galactose (2) (25.0 g, 64.1 mmol) was added 30% HBr/AcOH (50 mL), and the mixture was stirred for 3 h at room temperature. The reaction mixture was azeotropically evaporated with toluene under reduced pressure to afford a bromide 3. Without purification, it was dissolved in 1,2-dichloroethane (200 mL) and triethylamine (18.0 mL, 129 mmol), MeOH (2.74 mL, 67.6 mmol), and n-tetrabutylammonium bromide (10.3 g, 31.9 mmol) were added. The mixture was stirred for 16 h at 45 °C, and the precipitated salt was removed by filtration. The filtrate was washed with brine and concentrated in vacuo. The residue was chromatographed over Si gel. Elution with n-hexane/EtOAc (9:1, 8:2, and 7:3) gave 22.1 g of 4 (61.0 mmol, 95% from 2): ¹H NMR (300 MHz, CDCl₃) δ 5.78 (1H, d, J = 4.8 Hz), 5.40 (1H, dd, J = 2.9, 2.7 Hz), 5.03 (1H, dd, J = 6.8, 3.3 Hz), 4.29(2H, m), 4.10 (2H, m), 3.27 (3H, s), 2.08 (3H, s), 2.04 (3H, s), 2.03 (3H, s), 1.64 (3H, s).

3,4,6-Tri-*O***-benzyl-1,2-***O***-(1-methoxyethylidene)**- α -**D**-galactopyranose (5). To a stirred solution of **4** (22.0 g, 60.8 mmol) in toluene (60 mL) were added powdered KOH (35.0 g, 620 mmol) and benzyl chloride (48.0 mL, 417 mmol). The mixture was gradually heated over 1 h and refluxed for 1.5 h. It was then diluted with toluene and washed with water and brine. The toluene solution was concentrated in vacuo and chromatographed over Si gel. Elution with *n*-hexane/EtOAc (8:2) gave 19.1 g of **5** (37.5 mmol, 62%): ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.20 (15H, m), 5.73 (1H, d, J = 4.5 Hz), 4.90 (1H, d, J = 11.5 Hz), 4.79 (1H, d, J = 12.2 Hz), 4.66 (1H, d, J = 11.9 Hz), 4.42 (1H, d, J = 11.9 Hz), 4.03–3.98 (2H, m), 3.63–3.58 (3H, m), 3.25 (3H, s), 1.56 (3H, s).

3,4,6-Tri-*O***-benzyl-D-galactopyranose (6)**. To a stirred solution of **5** (19.0 g, 37.5 mmol) in 1,4-dioxane (280 mL) was added 1 M H₂SO₄ (80 mL), and the mixture was refluxed (110 °C) for 2 h. The reaction mixture was cooled to room temperature, then solid NaHCO₃ was carefully added until the mixture was neutralized; stirring was continued for 30 min.

The precipitated salts were removed by filtration, and the filtrate was evaporated to dryness. The resulting residue was partitioned between CHCl₃ and water; the organic layer was washed with brine and concentrated in vacuo to afford 15.5 g of **6** (34.4 mmol, 92%): ¹H NMR of the α -anomer (300 MHz, CDCl₃) δ 7.36–7.20 (15H, m), 5.27 (1H, d, J = 4.0 Hz), 4.84 (1H, d, J = 11.6 Hz), 4.69 (1H, d, J = 11.9 Hz), 4.62 (1H, d, J = 11.9 Hz), 4.62 (1H, d, J = 11.6 Hz), 4.52 (1H, d, J = 11.6 Hz), 4.46 (1H, d, J = 11.9 Hz), 4.38 (1H, d, J = 11.9 Hz), 4.10 (1H, dd, J = 10.0, 4.0 Hz), 3.88 (1H, m), 3.68 (1H, dd, J = 9.8, 2.8 Hz), 3.52 (1H, dd, J = 9.2, 6.7 Hz), 3.42 (1H, dd, J = 9.5, 6.1 Hz).

(2.S,3.S,4.R)-2,3,5-Tribenzyloxy-4-hydroxypentanal (7). Sodium metaperiodate (7.2 g, 33.6 mmol) was added to a solution of diol **6** (7.6 g, 16.9 mmol) in EtOH/H₂O (4:1, 100 mL), and the mixture stirred for 17 h at room temperature. The reaction mixture was concentrated and partitioned between ether and water. The organic layer was washed with brine and dried (Na₂SO₄), and the solvent evaporated to give an oil **7** (7.3 g), which was used in the next step without further purification.

(2R,3R,4R)-1,3,4-Tribenzyloxy-5-pentadecene-2-ol (8a). A suspension of decyltriphenylphosphonium bromide (18.0 g, 37.2 mmol) in THF (80 mL) was stirred at 0 °C, while 16.0 mL of *n*-BuLi (1.6 M solution in *n*-hexane) was added dropwise. The resulting solution was stirred for 30 min, and then a solution of the aldehyde 7 (4.6 g, 10.7 mmol) in THF (20 mL) was added. The mixture was warmed to room temperature and stirred for 1 h. The reaction was quenched by adding water and extracted with ether. The ether layer was dried with Na₂-SO₄ and concentrated in vacuo. SiO₂ chromatography with n-hexane/EtOAc (8:2) gave the alkene 8a (4.41 g, 8.1 mmol, 76%) as a pale yellow oil: ¹H NMR of the 5Z-isomer (300 MHz, $CDCl_3$) δ 7.35–7.20 (15H, m), 5.72 (1H, m), 5.45 (1H, dd, J= 11.0, 10.9 Hz), 4.68 (1H, d, J = 11.3 Hz), 4.60 (1H, d, J = 11.7 Hz), 4.50-4.45 (3H, m), 4.40 (1H, dd, J = 9.5, 5.2 Hz), 4.33 (1H, d, J = 11.8 Hz), 4.07 (1H, td, J = 5.9, 2.9 Hz), 3.56 (1H, td, J = 5.9, 2.9 Hz)), 3.56 (1H, td, J = 5.9, 2.9 Hz)))dd, J = 5.6, 2.9 Hz), 3.51 (2H, d, J = 5.9 Hz), 2.81 (1H, brs), 1.93 (2H, m), 1.30-1.20 (14H, m), 0.88 (3H, t, J = 7.0 Hz).

(2R,3R,4R)-1,3,4-Tribenzyloxy-5-hexadecene-2-ol (8b). A suspension of undecyltriphenylphosphonium bromide (16.5 g, 33.2 mmol) in THF (70 mL) was stirred at 0 °C, while 18.0 mL of *n*-BuLi (1.6 M solution in *n*-hexane) was added dropwise. The resulting solution was stirred for 10 min, and then a solution of the aldehyde 7 (4.6 g, 10.7 mmol) in THF (15 mL) was added. The mixture was warmed to room temperature and stirred for 30 min. The reaction was quenched by adding water and then extracted with ether. The ether layer was dried with Na₂SO₄ and concentrated in vacuo. SiO₂ chromatography with *n*-hexane/EtOAc (8:2) gave the alkene **8b** (4.15 g, 7.43 mmol, 69%) as a pale yellow oil: ¹H NMR of the 5Z-isomer (300 MHz, CDCl₃) δ 7.35–7.20 (15H, m), 5.71 (1H, dt, J = 11.0, 7.0 Hz), 5.44 (1H, dd, J = 11.0, 11.0 Hz), 4.66 (1H, d, J = 11.3 Hz), 4.61 (1H, d, J = 9.5 Hz), 4.50–4.45 (3H, m), 4.42 (1H, dd, J =10.3, 5.0 Hz), 4.33 (1H, d, J = 11.8 Hz), 4.06 (1H, td, J = 5.9, 2.9 Hz), 3.54 (1H, dd, J = 5.5, 2.9 Hz), 3.50 (2H, d, J = 5.9

Hz), 1.94 (1H, ddt, J = 13.5, 7.0, 7.0 Hz), 1.87 (1H, ddt, J = 13.5, 7.0, 7.0 Hz), 1.30–1.20 (16H, m), 0.86 (3H, t, J = 7.0 Hz).

(2R,3R,4R)-1,3,4-Tribenzyloxy-2-methanesulfonyloxy-5-pentadecene (9a). A solution of 8a (4.0 g, 7.17 mmol) in pyridine (10.0 mL) was ice-cooled, and then MsCl (1.20 mL, 15.5 mmol) was added. The mixture was stirred for 7 h at room temperature, then toluene was added, and the solution was concentrated in vacuo. The resulting residue was dissolved in ether, and the solution was washed successively with water and brine. The organic layer was dried over Na₂SO₄ and evaporated to dryness, and the residue was chromatographed over Si gel. Elution with n-hexane/EtOAc (9:1) gave the O-mesylate 9a (4.37 g, 6.87 mmol, 96%): ¹H NMR of the 5Zisomer (500 MHz, CDCl₃) & 7.35-7.20 (15H, m), 5.77 (1H, dt, J = 11.2, 7.3 Hz), 5.46 (1H, t, J = 11.2 Hz), 5.03 (1H, ddd, J = 7.1, 4.0, 4.0 Hz), 4.73 (1H, d, J = 11.0 Hz), 4.51-4.35 (6H, m), 3.74 (1H, dd, J = 11.0, 3.7 Hz), 2.90 (3H, s), 2.04 (1H, ddt, J = 14.5, 7.3, 7.3 Hz), 1.94 (1H, ddt, J = 14.5, 7.3, 7.3 Hz), 1.35-1.20 (16H, m), 0.84 (3H, t, J = 7.0 Hz).

(2R,3R,4R)-2-Methanesulfonyloxy-1,3,4-pentadecanetriol (10a). To an ice-cooled solution of 20a (4.0 g, 7.35 mmol) in pyridine (15.0 mL) was added MsCl (1.15 mL, 14.9 mmol). The mixture was stirred for 15 h at room temperature, then toluene was added, and the solution was concentrated in vacuo. The resulting residue was dissolved in ether, and the solution was washed successively with water and brine. The organic layer was dried over Na₂SO₄ and evaporated to dryness, and the residue was chromatographed over Si gel. Elution with n-hexane/EtOAc (9:1, 8:2) afforded the O-mesylate 9a (4.66 g, quant.). Then the O-mesylate 9a (2.26 g, 3.63 mmol) was dissolved in EtOH (30 mL), and 2.30 g of 10% palladium on carbon was added. Hydrogen gas was introduced to the reaction flask and the solution vigorously stirred for 20 h (room temperature, 1 atm). The reaction mixture was filtered through a pad of Celite (with CHCl₃ rinse), and the filtrate was concentrated to give 1.24 g (3.50 mmol, 96%) of 10a as a white powder: ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 4.89 (1H, ddd, J = 6.5, 5.3, 2.1 Hz), 3.90-3.75 (2H, m), 3.53-3.48(1H, m), 3.46 (1H, dd, J = 8.5, 2.1 Hz), 3.14 (3H, s), 1.74 (2H, m), 1.52 (2H, m), 1.40–1.20 (16H, m), 0.84 (3H, t, J=6.7 Hz).

(2*R*,3*R*,4*R*)-2-Methanesulfonyloxy-1,3,4-hexadecanetriol (10b). To a solution of 9b (1.35 g, 2.12 mmol) in EtOH (30 mL) was added 10% palladium on carbon (1.36 g). The mixture was stirred for 22 h under a hydrogen atmosphere at room temperature (1 atm). The Pd/C was then removed by filtration through a pad of Celite, and the filtrate was concentrated to give the triol **10b** as a white powder (1.24 g, 3.50 mmol, 89%): ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 4.90 (1H, ddd, J = 6.6, 5.3, 2.0 Hz), 3.86 (1H, dd, J = 12.2, 6.5Hz), 3.80 (1H, dd, J = 12.1, 4.9 Hz), 3.52 (1H, ddd, J = 8.4, 8.4, 2.3 Hz), 3.46 (1H, dd, J = 8.4, 1.9 Hz), 3.15 (3H, s), 1.75 (2H, m), 1.52 (2H, m), 1.40–1.20 (18H, m), 0.84 (3H, t, J =6.7 Hz).

(2.5,3.5,4.R)-2-Azido-1,3,4-pentadecanetriol (11a). Sodium azide (600 mg, 9.2 mmol) was added to a solution of the mesylate **10a** (609 mg, 1.72 mmol) in DMF (20 mL) at room temperature. The mixture was slowly heated to 100 °C over 30 min and then stirred for 3 h. The mixture was diluted with EtOAc and washed with water and brine. The organic layer was dried (Na₂SO₄) and concentrated in vacuo, and remaining DMF was removed by lyophilization. The resulting residue was chromatographed over Si gel with CHCl₃/MeOH (97:3, 96:4) to yield 425 mg (1.41 mmol, 82%) of the azide **11a**: ¹H NMR (600 MHz, CDCl₃/CD₃OD, 1:1) δ 3.90 (1H, dd, J = 12.0, 3.6 Hz), 3.74 (2H, m), 3.54 (2H, m), 1.62 (2H, m), 1.56 (2H, m), 1.40–1.20 (16H, m), 0.85 (3H, t, J = 7.0 Hz).

(2.5,3.5,4.R)-2-Azido-1,3,4-hexadecanetriol (11b). Sodium azide (200 mg, 3.08 mmol) was added to a solution of the mesylate **10b** (300 mg, 0.82 mmol) in DMF (12 mL) at room temperature. It was slowly heated to 100 °C over 30 min and then stirred for 3 h. The reaction mixture was partitioned between EtOAc and H₂O. The organic layer was dried (Na₂-SO₄) and concentrated in vacuo. Chromatography (SiO₂)

eluting with CHCl₃/MeOH (95:5) gave 191 mg (0.61 mmol, 74%) of the azide **11b**: ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 3.90 (1H, dd, J = 11.9 and 3.4 Hz), 3.75 (2H, m), 3.55 (2H, m), 1.65–1.47 (4H, m), 1.40–1.20 (18H, m), 0.84 (3H, t, J = 6.9 Hz).

(2S,3S,4R)-2-Amino-1,3,4-pentadecanetriol (12a). To a solution of the azide 11a (500 mg, 1.66 mmol) in EtOH (20 mL) was added 10% palladium on carbon (500 mg). The reaction flask was flushed with hydrogen gas and the solution vigorously stirred for 20 h (room temperature, 1 atm). The mixture was then filtered through a pad of Celite (with CHCl₃/ MeOH rinse), the solvent concentrated, and the residue chromatographed over Si gel under basic conditions. The column was eluted with CHCl₃/MeOH/H₂O/25% NH₄OH (8:2: 0.15:0.05), and the eluate was concentrated, diluted with water, and lyophilized to afford the C₁₅ phytosphingosine **12a** (433 mg, 1.57 mmol, 95%): ¹H NMR (300 MHz, CDCl₃) δ 3.72 (1H, dd, J = 11.1, 4.3 Hz), 3.58 (1H, dd, J = 11.1, 6.2 Hz),3.51 (1H, m), 3.32 (1H, dd, J = 7.7, 5.5 Hz), 2.94 (1H, dd, J = 10.1, 5.5 Hz), 1.70 (2H, m), 1.53 (2H, m), 1.40-1.20 (16H, m), 0.85 (3H, t, J = 6.9 Hz).

(2.5,3.5,4.R)-2-Amino-1,3,4-hexadecanetriol (12b). To a solution of the azide 11b (100 mg, 0.32 mmol) in EtOH (11 mL) was added 10% palladium on carbon (110 mg). The reaction flask was flushed with hydrogen gas and the solution vigorously stirred for 20 h (room temperature, 1 atm). The mixture was then filtered through a pad of Celite (with CHCl₃/MeOH rinse), the solvent concentrated, and the residue chromatographed over Si gel. Elution with CHCl₃/MeOH/H₂O (8:2:0.2, 7:3:0.5) gave the C₁₆ phytosphingosine12b (51.0 mg, 0.18 mmol, 56%): ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 3.87 (1H, dd, J = 11.8, 4.0 Hz), 3.75 (1H, dd, J = 11.8, 7.5 Hz), 3.62 (1H, m), 3.52 (1H, dd, J = 8, 4.4 Hz), 3.45 (1H, dd, J = 7.6, 3.7 Hz), 1.73 (2H, m), 1.49 (2H, m), 1.40–1.20 (18H, m), 0.83 (3H, t, J = 7.0 Hz).

9-Bromononanal (14). To a stirred solution of 9-bromononanol (13) (4.9 g, 22.0 mmol) in CH_2Cl_2 (30 mL) was added pyridinium chlorochromate (10.0 g, 46.4 mmol). Stirring was continued for 2 h at room temperature, and then ether (10 mL) and Na_2SO_4 were added to the solution. The suspension was filtered and rinsed with ether, and the solvent concentrated in vacuo. The residue was chromatographed over Si gel. Elution with *n*-hexane/EtOAc (96:4, 9:1) gave the aldehyde **14** (4.3 g, 19.5 mmol, 89%): ¹H NMR (300 MHz, CDCl₃) δ 9.74 (1H, t, J = 1.8 Hz), 3.38 (2H, t, J = 7.0 Hz), 2.40 (2H, brt, J = 7.0 Hz), 1.82 (2H, quint, J = 7.0 Hz), 1.35–1.25 (6H, m).

12-Bromo-2-methyl-3-dodecene (15). To an ice-cooled suspension of isobutyltriphenylphosphonium bromide (10.0 g, 25.0 mmol) in THF (50 mL) was added 14.0 mL of n-BuLi (1.6 M solution in *n*-hexane). The resulting solution was stirred for 30 min at 0 °C, and then a solution of 8-bromooctanal (4.1 g, 18.5 mmol) in THF (30 mL) was added. The mixture was warmed to room temperature and stirring was continued for 2 h. The reaction was quenched by addition of water, and the mixture extracted with ether. The combined ethereal fraction was washed with brine, dried over MgSO₄, and concentrated. Chromatography on Si gel with *n*-hexane gave the alkene **15** as a colorless oil (3.8 g, 14.4 mmol, 78%): ¹H NMR of the 3*Z*isomer (300 MHz, CDCl₃) δ 5.20 (1H, dt, J = 11.0, 6.9 Hz), 5.16 (1H, ddt, J = 10.7, 10.7, 1.2 Hz), 3.38 (2H, t, J = 6.7 Hz), 2.56 (1H, m), 2.00 (2H, dt, J = 7.0, 7.0 Hz), 1.83 (2H, quint, J = 6.9 Hz), 1.42 (2H, m), 1.40–1.20 (8H, m), 0.92 (6H, d, J =6.7 Hz)

11-Methyl-9-dodecenyltriphenylphosphonium Bromide (16). The mixture of the bromide **15** (3.6 g, 16.2 mmol) and triphenylphosphine (4.3 g, 16.4 mmol) was heated at 120 °C for 2 days to give **16**. In the same manner, this reaction was repeated twice for each of the next Wittig reactions. The resulting viscous materials were used in the next step without further purification.

1-Bromo-19-methyleicosane (18a). To a suspension of the phosphonium salt **16** (prepared from 3.6 g of **15**) in THF (20

mL) was added 8.5 mL of n-BuLi (1.6 M solution in n-hexane) at 0 °C. The resulting solution was stirred for 30 min at 0 °C, and then a solution of 8-bromoctanal (3.4 g, 16.4 mmol) in THF (15 mL) was added. The mixture was warmed to room temperature, and stirring was continued for 1 h. The reaction was quenched by the addition of water, and the mixture extracted with ether. The combined ethereal fraction was washed with brine, dried over MgSO₄, and concentrated. Chromatography over Si gel with *n*-hexane afforded the *E*,*Z*mixture of the 1-bromo-19-methyl-8,17-eicosadiene 17a as a colorless oil (2.7 g, 7.3 mmol, 45%). The ¹H NMR spectrum of 17a exhibited additional signals for 1-bromo-19-methyl-8,16eicosadiene and 1-bromo-19-methyl-8,18-eicosandiene probably produced through migration of the double bond, which were not separated, but were used as such in the next reaction. The mixture of 17a (2.7 g, 7.3 mmol) and 10% palladium on carbon (2.7 g) in 30 mL of EtOH was stirred under a hydrogen atmosphere for 20 h (room temperature, 1 atm). The reaction mixture was filtered through a pad of Celite (with CHCl₃ rinse), and the filtrate was evaporated to dryness. Chromatography over Si gel, eluting with n-hexane, yielded the bromoalkane 18a (2.0 g, 5.4 mmol, 74%) as a colorless oil: 1H NMR (300 MHz, CDCl₃) δ 3.39 (2H, t, J = 6.8 Hz), 1.84 (2H, quint, J = 7.3 Hz), 1.50 (1H, m), 1.41 (2H, m), 1.35-1.20 (28H, \hat{m}), 1.20–1.15 (2H, m), 0.85 (6H, d, J = 6.7 Hz).

1-Bromo-20-methylheneicosane (18b). Following the same procedure as described above for the preparation of **18a**, 9-bromononanal (2.0 g, 9.0 mmol) gave **18b** (1.1 g, 2.8 mmol, 31%): ¹H NMR (300 MHz, CDCl₃) δ 3.39 (2H, t, J = 7.1 Hz), 1.83 (2H, quint, J = 6.7 Hz), 1.49 (1H, m), 1.40 (2H, m), 1.35 – 1.20 (30H, m), 1.13 (2H, m), 0.84 (6H, d, J = 6.8 Hz).

1-Bromo-21-methyldocosane (18c). Following the same procedure as described above for the preparation of **18a**, 10-bromodecanal (3.7 g, 15.7 mmol) afforded **18c** (4.0 g, 10.0 mmol, 64%): ¹H NMR (300 MHz, CDCl₃) δ 3.40 (2H, t, J = 7.0 Hz), 1.83 (2H, quint, J = 6.8 Hz), 1.50 (1H, m), 1.40 (2H, m), 1.35–1.20 (32H, m), 1.15 (2H, m), 0.85 (6H, d, J = 6.8 Hz).

2-Hydroxy-21-methyldocosanoic acid (20a). To a suspension of diethyl acetamidomalonate (5.0 g, 23.0 mmol) in EtOH (10.0 mL) was added 9.0 mL of sodium ethoxide (2.4 M solution in EtOH, prepared from Na and EtOH). The mixture was stirred for 30 min at room temperature. Then, 7.0 mL (6.0 mmol) of the resulting solution was added to a solution of the bromide 18a (935 mg, 2.49 mmol) in EtOH (5.0 mL), and the mixture was stirred and heated under reflux for 10 h. The reaction mixture was concentrated, diluted with water, and extracted with ether. The ether extract was concentrated in vacuo. The residue was dissolved in concentrated HCl (7.5 mL), water (2.5 mL), and 1,4-dioxane (5.0 mL), and the resulting mixture was stirred and heated under reflux for 12 h. The reaction mixture was lyophilized to give 500 mg (1.36 mmol) of amino acid 19a as a white powder. Part of the resulting amino acid (91.0 mg, 0.26 mmol) was mixed with 2 N H₂SO₄ (5.0 mL) and 1,4-dioxane (5.0 mL). A solution of sodium nitrite (75.0 mg, 1.09 mmol) in water (1.0 mL) was added to the suspension, which was stirred for 1 h at 80 °C. The resulting solution was diluted with ether, washed with water, dried (MgSO₄), and concentrated. Chromatography on Si gel with n-hexane/EtOAc/AcOH (5:5:0.2) afforded the 2-hydroxy acid 20a (56.0 mg, 0.15 mmol, 32% from 18a): ¹H NMR (300 MHz, $CDCl_3/CD_3OD$, 1:1) δ 4.09 (1H, dd, J = 7.6, 4.5 Hz), 1.80-1.50 (2H, m), 1.50-1.20 (33H, m), 1.12 (2H, m), 0.82 (6H, d, J = 6.7 Hz).

2-Hydroxy-22-methyltricosanoic acid (20b). Following the same procedure as described above for the preparation of **20a**, the bromoalkane **18b** was converted to the 2-hydroxy acid **20b** (18%): ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 4.09 (1H, dd, J = 7.6, 4.5 Hz), 1.80–1.50 (2H, m), 1.50–1.20 (35H, m), 1.12 (2H, m), 0.82 (6H, d, J = 6.5 Hz).

2-Hydroxy-23-methyltetracosanoic acid (20c). Following the same procedure as described above for the preparation of **20a**, the bromoalkane **18c** was converted to the 2-hydroxy acid **20c** (27%); ¹H NMR (300 MHz, CDCl₃/CD₃OD, 1:1) δ 4.09

(1H, dd, J = 7.6, 4.5 Hz), 1.80–1.50 (2H, m), 1.50–1.20 (37H, m), 1.12 (2H, m), 0.82 (6H, d, J = 6.5 Hz).

2-Acetoxy-21-methyldocosanoic acid (21a). Acetic anhydride (4.0 mL) was added to a solution of the alcohol **20a** (117 mg, 0.32 mmol) in pyridine (4.0 mL). The mixture was stirred for 13 h at room temperature. It was then diluted with ice water and extracted with ether. The ethereal extract was washed with brine, dried (MgSO₄), and concentrated. The residue was chromatographed over Si gel. Elution with *n*-hexane/EtOAc/AcOH (5:5:0.2) gave the acetate **21a** as a white solid (102 mg, 0.25 mmol, 77%): ¹H NMR (600 MHz, CDCl₃) δ 4.94 (1H, dd, J = 7.3, 5.0 Hz), 2.07 (3H, s), 1.79 (2H, m), 1.44 (2H, m), 1.36 (2H m), 1.40–1.20 (30H, m), 1.07 (2H, m), 0.79 (6H, d, J = 6.5 Hz).

2-Acetoxy-22-methyltricosanoic acid (21b). In the same manner as described above for the preparation of **21a**, the alcohol **20b** (109 mg, 0.28 mmol) was converted to 105 mg (0.25 mmol, 88%) of **21b**: ¹H NMR (600 MHz, CDCl₃) δ 4.99 (1H, t, J = 6.3 Hz), 2.12 (3H, s), 1.84 (2H, m), 1.49 (2H, m), 1.41 (2H m), 1.40–1.20 (30H, m), 1.13 (2H, m), 0.84 (6H, d, J = 6.9 Hz).

2-Acetoxy-23-methyltetracosanoic acid (21c). In the same manner as described above for the preparation of **21a**, the alcohol **20c** (180 mg, 0.45 mmol) was converted to 168 mg (0.38 mmol, 84%) of **21c**: ¹H NMR (600 MHz, CDCl₃) δ 4.98 (1H, dd, J = 7.3, 6.4 Hz), 2.12 (3H, s), 1.84 (2H, m), 1.49 (2H, m), 1.40 (2H, m), 1.40–1.20 (30H, m), 1.13 (2H, m), 0.83 (6H, d, J = 6.5 Hz).

Synthesis of the Ceramide Mixture (1a, 1b). The mixture of phytosphingosines 12a (25.0 mg, 0.091 mmol) and 12b (33.0 mg, 0.114 mmol), 2-acetoxy fatty acids 21a (30.0 mg, 0.073 mmol), 21b (30.0 mg, 0.070 mmol), and 21c (20.0 mg, 0.045 mmol), and HOBt (1-hydroxybendotriazole) (40.0 mg, 0.30 mmol) was diluted with benzene, evaporated, and lyophilized to remove water. The residue was dissolved in CH₂Cl₂ (5.0 mL) and added to WSCI [1-(3-dimethylaminopropyl)-3ethylcarbodiimide hydrochloride] (60.0 mg, 0.31 mmol). The mixture was stirred for 20 h at room temeperature, then diluted with CHCl₃, washed with water, dried (NaSO₄), and concentrated. To the residue were added MeOH (9.0 mL), THF (3.0 mL), and K₂CO₃ (90 mg), and the mixture was stirred for 2.5 h at room temperature. The reaction mixture was concentrated in vacuo and chromatographed over Si gel. Elution with CHCl₃/MeOH (97:3, 96:4) furnished 2'S (1 b, 45.0 mg) and 2'R (1 a, 36.0 mg) ceramide mixtures. 1a: ¹H NMR (600 MHz, CDCl₃/CD₃OD, 1:1) δ 4.08 (1H, ddd, J = 4.8, 4.6, 3.8 Hz), 4.00 (1H, dd, J = 8.1, 3.8 Hz), 3.76 (1H, dd, J = 11.2, 4.6 Hz), 3.70 (1H, dd, J = 11.3, 4.8 Hz), 3.51 (2H, m), 1.74 (1H, m), 1.64 (1H, m), 1.60-1.15 (brm), 1.12 (2H, m), 0.84 (3H, t, J = 6.9Hz), 0.82 (6H, d, J = 6.9 Hz); ¹³C NMR (150 MHz, CDCl₃/CD₃-OD, 1:1) & 176.4s, 75.9d, 72.9d, 72.4d, 61.6t, 52.1d, 39.6t, 35.1t, 33.2t, 32.5t, 30.4t, 30.2 (18-21C), 29.9t, 28.5t, 27.9t, 26.4t, 25.6d, 23.2q (2C), 22.9t, 14.3q; HRFABMS for C₁₅/C₂₃ m/z (M + H)⁺ 628.5873 (calcd for $C_{38}H_{78}NO_5 \Delta$ -0.7 mmu); C_{16}/C_{23} m/z (M + H)⁺ 642.6085 (calcd for C₃₉H₈₀NO₅ Δ +4.8 mmu); $C_{15}/C_{25} m/z (M + H)^+ 656.6244$ (calcd for $C_{40}H_{82}NO_5 \Delta + 5.1$ mmu); C₁₆/C₂₅ m/z (M + H)⁺ 670.6400 (calcd for C₄₁H₈₄NO₅ Δ +5.0 mmu). 1b: ¹H NMR (600 MHz, CDCl₃/CD₃OD, 1:1) δ 4.09 (1H, ddd, J = 5.0, 4.6, 3.8 Hz), 4.00 (1H, dd, J = 8.1, 3.8 Hz),3.76 (1H, dd, J = 11.2, 4.6 Hz), 3.70 (1H, dd, J = 11.2, 5.0 Hz), 3.52 (2H, m), 1.76 (1H, m), 1.64 (1H, m), 1.60-1.15 (brm), 1.12 (2H, m), 0.84 (3H, t, J = 6.9 Hz), 0.82 (6H, d, J = 6.6Hz).

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Supporting Information Available: ¹H NMR spectra and HPLC profiles of the natural and synthetic ceramide mixtures. This material is available free of charge via the Internet at http://pubs.acs.org.

Sex Pheromones of the Hair Crab Erimacrus

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- (19) ¹³C NMR data for the natural ceramide mixture (150 MHz, CDCl₃/ CD₃OD, 1:1) δ 176.5s, 75.7d, 72.7d, 72.5d, 61.5t, 52.0d, 34.8t, 33.0t, 30.0t, 29.5d, 24.0t, 22.7q(2C), 14.2q.

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